

REVIEW ARTICLE

A review of genetic epidemiology of head and neck cancer related to polymorphisms in metabolic genes, cell cycle control and alcohol metabolism

Revisione narrativa sulla epidemiologia genetica dei polimorfismi, dei geni metabolici, dei geni del ciclo cellulare e dei geni del metabolismo dell'alcool, implicati nel rischio del cancro testa-collo

G. CADONI¹, S. BOCCIA^{2,3}, L. PETRELLI¹, P. DI GIANNANTONIO², D. ARZANI², A. GIORGIO¹, E. DE FEO², M. PANDOLFINI¹, P. GALLI², G. PALUDETTI¹, G. RICCIARDI²

¹ Institute of Otorhinolaryngology, Università Cattolica del Sacro Cuore, Rome, Italy; ² Institute of Hygiene, Università Cattolica del Sacro Cuore, Rome, Italy; ³ IRCCS, San Raffaele Pisana, Rome, Italy

SUMMARY

The purpose of this report is to review the relationship between genetic polymorphisms involved in carcinogen metabolism, alcohol metabolism and cell-cycle control with the risk of head and neck cancer. The review was performed on available studies on genetic polymorphisms and head and neck cancer (HNC) published in PubMed up to September 2011. 246 primary articles and 7 meta-analyses were published. Among these, a statistically significant association was reported for *glutathione S-transferases (GSTM1)*, *glutathione S-transferases (GSTT1)* and *human microsomal epoxide hydrolase (EPHX1)* genes. An increased risk for HNC was also associated reported for *P53* codon 72 Pro/Pro, *ALDH2* and three variants of the *ADH* gene: *ADH1B (rs1229984)*, *ADH7 (rs1573496)* and *ADH1C (rs698)*.

KEY WORDS: Head and neck cancer • Genetic susceptibility • Genetic polymorphisms

RIASSUNTO

Lo scopo di questo lavoro è stato valutare la relazione esistente tra i polimorfismi dei geni coinvolti nella carcinogenesi testa-collo, nel metabolismo dell'alcool e nel controllo del ciclo cellulare e il rischio di sviluppare un tumore della testa e del collo (HNC). La revisione è stata effettuata su studi relativi ai polimorfismi genetici e al cancro testa collo (HNC) pubblicati su PubMed fino al settembre 2011. I risultati hanno mostrato l'esistenza di 246 articoli primari e 7 meta-analisi. Tra queste, una associazione statisticamente significativa è stata segnalata per la glutathione S-transferasi (GSTM1), glutathione S-transferasi (GSTT1) e Human microsomal epoxide hydrolase (EPHX1) geni. Un aumentato rischio di sviluppo del tumore testa collo è stato inoltre associato alla presenza della variante di P53 codone 72 Pro/Pro, di ALDH2, e delle tre varianti del gene ADH: ADH1B (rs1229984), ADH7 (rs1573496) e ADH1C (rs698).

PAROLE CHIAVE: Tumore testa-collo • Suscettibilità genetica • Polimorfismi genetici

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Introduction

Head and neck cancers (HNC), including oral cavity, oropharynx, hypopharynx and larynx are among the most common types of cancer and represent a major health problem¹. There are approximately 540,000 new cases and 271,000 deaths annually worldwide for a mortality of approximately 50%. HNC represent approximately 3% of all cancers in the United States², but are much more prevalent in some parts of South America (Brazil, Uruguay and Argentina)¹. Development of HNC is a multifactorial process associated with a variety of risk factors. Tobacco and alcohol consumption are established risk factors for HNC (synergistic in persons who both smoke tobacco and drink alcohol), even though other factors may affect risk^{3,4}. In

developing countries, other factors, like betel and tobacco chewing, human papillomavirus infection or drinking hot beverages, also play an important role¹. In recent years, evidence has accumulated to support the hypothesis that diet may also play an important aetiological role in development of disease, and 10-15% of squamous cell carcinoma of the head and neck (SCCHN) cases in Europe are associated with a low intake of fruit and vegetables⁵⁻⁸. A family history of cancer is another important risk factor for HNC⁹, which implies that genetics contributes to HNC susceptibility¹⁰.

Materials and methods

Herein, we review the published studies concerning genetic susceptibility to HNC. A variety of genes are associ-

ated with HNC carcinogenesis, including those involved in carcinogen metabolism, alcohol metabolism, folate metabolism, DNA repair, cell-cycle control and oncogenes². We have focused our attention on genes involved in carcinogen metabolism, alcohol metabolism and cell-cycle control.

A PubMed search was performed until to the end of September 2011. The following terms were used: head and neck and cancer (with both synonymous and plural forms), as well as the truncated words genetic*, allel*, or polymorphi*.

Results

The literature search identified the below list of genes:

- *Cytochrome P450 2E1 (CYP2E1)*;
- *Cytochrome P450 1A1 (CYP1A1)*;
- *Glutathione S-transferase (GSTM1)*;
- *Glutathione S-transferase (GSTT1)*;
- *Glutathione S-transferase (GSTP1)*;
- *N-acetyltransferase 2 (NAT2)*;
- *Human microsomal epoxide hydrolase (EPHX1)*;
- *Aldehyde dehydrogenase 2 (ALDH2)*;
- *Alcohol dehydrogenase isoenzymes (ADH)*;
- *X-ray repair cross complementary 1 (XRCC1)*;
- *Xeroderma pigmentosum complementary group D (XPD)*;
- *Cyclin D1 (CCND1)*;
- *P53 tumour suppressor*;
- *P73*.

Tables I and II report, respectively, a summary of primary studies of genetic polymorphisms in genes involved in

Table I. Summary of primary studies of genetic polymorphisms and HNC risk.

Gene and polymorphism	No. of included studies
Cytochrome P450 2E1 (CYP2E1)	15
Cytochrome P450 1A1 (CYP1A1) codon 462	9
Glutathione S-transferase (GSTM1)	20
Glutathione S-transferase (GSTT1)	9
Glutathione S-transferase (GSTP1)	22
N-acetyltransferase 2 (NAT2)	11
Human microsomal epoxide hydrolase (EPHX1)	91
Aldehyde dehydrogenase 2 (ALDH2)	8
Alcohol dehydrogenase isoenzymes (ADH)	12
X-ray repair cross complementary 1 (XRCC1)	2
Xeroderma pigmentosum complementary group D (XPD)	6
Cyclin D1 (CCND1)	12
P53 tumour suppressor gene	26
P73	3

carcinogen metabolism, alcohol metabolism and cell-cycle control and the risk of HNC, and a summary of meta-analyses of genetic polymorphisms of the aforementioned gene classes and HNC risk.

Below we discuss the significance of each gene in the aetiology of HNC:

- *Cytochrome P450 2E1 (CYP2E1)*, a member of the cytochrome P-450 superfamily, is a naturally ethanol-inducible phase I enzyme. It is mainly involved in the metabolic activation of low molecular weight compounds such as nitrosamines and alcohol metabolism¹¹. The variant c2 allele, which contains a novel RsaI/PstI site in the 5'-flanking region of the *CYP2E1* gene, appears to be associated with decreased enzyme activity. Ten¹²⁻²¹ of the 15 studies¹²⁻²⁵ published before July 2007 suggested that the c1/c2 genotype of *CYP2E1* may increase risk for HNC compared with the c1/c1 genotype. Results of 6^{14 16 18 21 22 24} of 7 studies^{13 14 16 18 21 22 24} suggested that the c2/c2 genotype may increase risk for HNC².
- *Cytochrome P450 1A1 (CYP1A1)* is an important phase I enzyme that plays an essential role in the metabolic activation of major classes of pro-carcinogens such as benzopyrene, a prototypic polycyclic aromatic hydrocarbon²⁶. An Ile-Val substitution in codon 462 of *CYP1A1*, which is in the haem-binding region, results in a 2-fold increase in microsomal enzyme activity and, in Caucasians, is in complete linkage disequilibrium with the *CYP1A1* MspI polymorphism, which is also associated with increased catalytic activity. We identified some studies related to the association of *CYP1A1* with HNC risk published before July 2007, regarding the relation of the *CYP1A1* Ile-Val substitution at codon 462 to HNC. In 4 studies²⁷⁻³⁰, the risk for HNC in subjects with the Ile/Val and/or Val/Val genotypes was significantly higher than that for subjects with the Ile/Ile genotype, suggesting that the Val allele may be associated with increased risk for HNC. A meta-analysis of studies that examined the association of the *CYP1A1* Ile-Val substitution with risk for HNC revealed that the Ile/Val and Val/Val genotypes tend to increase HNC risk with an odds ratio (OR) [95% confidence interval (CI)] compared with Ile/Ile of 1.32 (0.95-1.82). Successive studies suggest that significant differences in the distribution of certain haplotypes of *CYPs* have been reported, and the prevalence of certain genotype combinations of *CYPs* and *GSTs* has indicated the importance of gene-gene interactions in HNSCC risk^{31 32}. The pooled data indicated that *CYP1A1* MspI polymorphism might be a risk factor for laryngeal cancer, particularly in Caucasians. For Asians, it might slightly increase the susceptibility to laryngeal cancer. However, the data failed to demonstrate a marked association between the *CYP1A1* exon 7 polymorphism and laryngeal cancer risk²⁶. The *CYP1A1* exon 7 polymorphism

Table II. Summary of previous meta-analyses of genetic polymorphisms and HNC risk.

Gene and polymorphism	Authors and year	No. of studies included	Associations studied	OR (95%CI)
Cytochrome P450 1A1 (CYP1A1) codon 462	Hashibe et al., 2003 ⁵⁰	12	Ile/Val+Val/Val vs. Ile/Ile	1.32 (0.95-1.82)
Glutathione S-transferase (GSTM1)	Hashibe et al., 2003 ⁵⁰	30	Null vs. Positive	1.23 (1.06-1.42)
	Tripathy & Roy, 2006 ⁵¹	30	Null vs. Positive	1.50 (1.21-1.87)
Glutathione S-transferase (GSTT1)	Hashibe et al., 2003 ⁵⁰	21	Null vs. Positive	1.17 (0.98-1.40)
Glutathione S-transferase (GSTP1)	Hashibe et al., 2003 ⁵⁰	9	Ile/Val+Val/Val vs. Ile/Ile	1.10 (0.92-1.31)
			Slow acetylators vs. cancer	0.99 (0.71-.38)
N-acetyltransferase 2 (NAT2)	Ying et al., 2011 ⁸⁰	7	Rapid acetylators vs. cancer	1.01 (0.72-.40)
			Y113H allele vs. cancer	0.86 (0.77-0.97)
Human microsomal epoxide hydrolase (EPHX1)	Li et al., 2011 ⁸³	82	H139R allele vs. cancer	1.05 (0.93-1.17)
			(RR) of H139R allele vs. cancer	1.34 (0.98-1.82)
XRCC1 codon 194	Hu et al., 2005 ¹⁴⁹	3	Arg/Trp+Trp/Trp vs. Arg/Arg	0.85 (0.59-1.23)
XRCC1 codon 399		4	Gln/Gln vs. Arg/Arg	1.13 (0.81-1.58)

was associated with oral and pharyngeal cancer only for ever smokers, when studied independently in the pooled analysis, although the *CYP1A1* MspI variant homozygote allele (m2/m2) was significantly associated with this cancer in both the meta-analysis and pooled analysis. When analyzing the complete genotype of *GSTM1* deletion and *CYP1A1* MspI polymorphism, the risk of oral and pharyngeal cancers seems to be higher for never smokers than for ever smokers. It should be highlighted that the results of the pooled analysis varied according to the type of controls considered, indicating that a selection bias might be present in some studies, and therefore the results should be considered with caution. There is no indication for population testing of these genes as risk factors for oral and pharyngeal cancer³³. Overall, Zhou et al. 2009 reported that variant genotypes of *CYP1A1* might not be risk factors for oral cancer.

- The glutathione S-transferases are a family of phase II xenobiotic metabolizing enzymes catalyzing the conjugation reactions of reactive intermediates of electrophilic compounds with cytosolic glutathione. Based on sequence similarities, human cytosolic glutathione S-transferases are mainly coded for at 5 loci: *GSTA* (*a*), *GSTT1* (*h*), *GSTM1* (*l*), *GSTP1* (*p*), and *GSTM3* (*c*). Polymorphisms in these genes, possibly by altering their expression and functional activities, may affect carcinogen activation/detoxification and DNA repair. Three alleles have been identified at the glutathione S-transferase M1 (*GSTM1*) locus: *GSTM1**0, *GSTM1**A, and *GSTM1**B. Two major alleles have been identified at the glutathione S-transferase T1 (*GSTT1*) locus: *GSTT1**1 and *GSTT1**0. Previous studies showed that a homozygous deletion (0/0), or null genotype, at either the *GSTM1* locus or the *GSTT1* locus resulted in

enzyme function loss³²⁻³⁴, and thus it was hypothesized to be related to the susceptibility to HNC.

- For HNCs since July 2007, 36 ORs from 58 studies of the null *GSTM1* genotype vs the positive genotype were > 1, suggesting that the null *GSTM1* genotype may be associated with increased risk for HNC. Sixteen^{17 35-49} of the studies showed a significantly higher risk for HNC in subjects with the null *GSTM1* genotype than in subjects with the positive genotype. Two meta-analyses^{50 51} of studies that examined the association of *GSTM1* with risk for HNC revealed that the null genotype significantly increases the risk with ORs (95%CI) of 1.23 (1.06-1.42) and 1.50 (1.21-1.87) compared with the positive genotype². Successive studies suggest that *GSTM1* has been reported to detoxify the bioreactive diol-epoxides of PAHs, which is important in environmental and occupational carcinogenesis. The data supported that *GSTM1* deficiency was associated with laryngeal cancer risk.
- We identified 23 ORs from 42 studies published before July 2007 of the null *GSTT1* genotype vs the positive genotype that were > 1, and 7 studies^{39 44 48 52-55} showing a significantly higher risk for HNC in subjects with the null genotype than with the positive genotype, suggesting that the null *GSTT1* genotype may be associated with increased risk for HNC. A meta-analysis⁵⁰ of studies that examined the association of *GSTT1* with risk of HNC revealed that the null genotype tends to increase HNC risk with ORs (95%CI) of 1.17 (0.98-1.40) compared with positive genotype².
- For *GSTP1*, four⁵⁶⁻⁵⁹ of the 21 studies^{14 47 56-74} showed a significantly higher risk for HNC in individuals with the Ile/Val and/or Val/Val genotypes than in those with the Ile/Ile genotype. No studies showed a significantly lower risk with the Ile/Val and/or Val/Val genotypes

than the Ile/Ile genotype. The 105 Val allele might be associated with an increased risk for HNC. One meta-analysis revealed that the Ile/Val and Val/Val genotypes tend to increase HNC risk with ORs (95%CI) of 1.10 (0.92-1.31) compared with the positive genotype^{3 50}. There have been no studies after July 2007 investigating the correlation between HNC and *GSTP1*.

- *Human arylamine N-acetyltransferases* play a key role in the metabolism of drugs and environmental chemicals and in the metabolic activation and detoxification of procarcinogens. Phenotyping analyses have revealed an association between *NAT* enzyme activities and the risk of developing several forms of cancer^{32 75}. The *NAT2* isoenzyme functions to both activate and deactivate arylamine and hydrazine drugs and carcinogens. Polymorphisms in this gene are responsible for the N-acetylation polymorphism in which human populations segregate into rapid, intermediate and slow acetylator phenotypes. Polymorphisms in *NAT2* are also associated with higher incidences of cancer and drug toxicity. A second *arylamine N-acetyltransferase gene (NAT1)* is located near *NAT2* (RefSeq, Jul 2008). For HNC, all 7 ORs^{14 23 64 76} for the slow *NAT2* genotype vs the rapid genotype were > 1, suggesting that the slow *NAT2* genotype may be associated with an increased risk for HNC². Ying XJ⁸⁰, in his meta-analysis published in 2011, claimed that there was overall lack of association between *NAT2* polymorphism and laryngeal cancer risk; however, *NAT2* slow acetylation may contribute to a risk factor for laryngeal cancer in Asians, but not in Caucasians. In 2008, Buch⁸¹ published results that demonstrated how fast *NAT2* acetylation was a risk factor for oral cancer. Demokan in 2010⁸² published that *NAT1* and *NAT2* gene combinations may influence the risk of developing head and neck cancer.
- *Human microsomal epoxide hydrolase (EPHX1)* plays an important role during xenobiotic detoxification of exogenous chemicals such as polycyclic aromatic hydrocarbons (PAHs), which are produced during the use of coal tar, coke, bitumen or during cigarette smoking^{32 83}. Two amino acid-altering polymorphisms, Tyr113His and His139Arg, have been identified in *EPHX1* and both are associated with alterations in mEH activity. The *EPHX1* His113 variant shows a 40% decrease in EH activity, whereas the *EPHX1* Arg139 variant shows 25% increased enzyme activity². A comprehensive systematic review of available studies published up to 2011, consisting of 91 studies, 84 (31,144 cases and 42,439 controls) for Tyr113His and 77 (28,496 cases and 38,506 controls) for His139Arg⁸³. Results of analysis of these two polymorphisms in different cancer types revealed that the low activity allele (H) of Y113H was highly associated with decreased risk of lung cancer (OR = 0.88, 95%CI = 0.80-0.96; p = 0.005) and HNC (OR = 0.86, 95%CI = 0.77-0.97; p = 0.014); the high activity allele (R) of H139R was significantly associated with increased risk of lung cancer (OR = 1.18, 95%CI = 1.04-1.33; p = 0.010), but not of HNC (OR = 1.05, 95%CI = 0.93-1.17, p = 0.447). However, the homozygous variant (RR) of H139R showed increased risk of HNC (OR = 1.34, 95%CI = 0.98-1.82, p = 0.065).
- *Aldehyde dehydrogenase 2 (ALDH2)* is a key gene in alcohol metabolism, and determines blood acetaldehyde concentrations after drinking¹⁰. A single point alteration in *ALDH2* results in the *ALDH2*2* allele. The protein encoded by *ALDH2*2* has a Glu to Lys substitution at residue 487, resulting in an inactive subunit and the inability to metabolize acetaldehyde. The *ALDH2*2* allele is rare in Western populations, but prevalent in East Asian populations. *ALDH2*2/*2* homozygotes have serum acetaldehyde levels that are 13 times higher and heterozygotes have levels 4 times higher than those in **1*1* homozygotes. Six studies^{13 40 84-87} reported a relation between *ALDH2* polymorphisms and risk for HNC, and all were conducted in Japanese populations. Four studies^{35 79 80 82} showed a significantly increased risk for HNC in **1/*2* heterozygotes compared with **1*1* homozygotes². Mc Kay et al. found that five genetic variants at three loci, 4q21, 4q23 and 12q24, were significantly associated with HNC risk¹⁰.
- *ADH isoenzymes*, which are primarily involved in ethanol oxidation, consist of subunits encoded by *ADH2* and *ADH3*. In contrast to *ADH2*, *ADH3* is highly polymorphic in Caucasians. Of the 2 allelic variants, the *ADH3*1* allele is associated with higher enzyme activity than the *ADH3*2* allele and occurs in Caucasians at frequencies of 55-63%. In 5^{19 37 88-90} of 8 studies^{19 37 88-93}, *ADH3*2/*1* heterozygotes showed decreased risk for HNC compared with **2/*2* homozygotes. However, in 6^{19 37 88 91-93} of 9 studies^{19 37 88-93}, *ADH3*1/*1* homozygotes showed increased risk for HNC². In accordance with Mc Kay and co-workers, three independent variants, *ADH1B* (rs1229984), *ADH7* (rs1573496) and *ADH1C* (rs698), have also been associated with HNC risk in European populations. The effects of these three variants were generally present for each HNC subsites, but more pronounced in oesophageal cancers and males. Strong heterogeneity was found with rs1229984 when stratifying by alcohol consumption.
- Polymorphisms in *X-ray repair cross complementing protein 1 (XRCC1)*, including Arg194Trp, Arg280His and Arg399Gln, have been described. Although the biochemical and biologic characteristics of the variants have not been determined, it has been reported that individuals with the *XRCC1* 399Gln variant show increased sister chromatid exchange after treatment with a tobacco-specific carcinogen². However, as already described by Gonzalez et al.⁹⁴ in 2002, the functional roles for these polymorphisms are unknown, even if

they are associated with high risk in non-smokers and in non-alcohol drinkers. The results for the relationship between *XRCC1* polymorphisms and HNC are inconsistent as highlighted in two meta-analyses published in 2005.

- *Xeroderma pigmentosum complementary group D (XPD)* has 2 functions: nucleotide excision repair and basal transcription as part of the transcription factor complex, TFIIH⁹⁵. Polymorphisms, such as 22,541AC and 35,931CA, have been identified. Individuals homozygous for the variant genotype of *XPD* have sub-optimal DNA repair capacity⁹⁶. Hiyama et al. in 2008 analyzed 4 studies⁹⁷⁻¹⁰⁰ of the genotype at nucleotide 22,541 and 5 studies⁹⁸⁻¹⁰² of the genotype at nucleotide 35,931 of *XPD*. According to these investigations, individuals homozygous for the variant genotype of *XPD* have suboptimal DNA repair capacity⁹⁶.
- The *CCND1* gene encodes a key cell cycle regulatory protein, cyclin D1, which regulates transition from G1 to the S phase during cell division. High activity of cyclin D1 leads to premature cell passage through the G1-S transition, resulting in propagation of unrepaired DNA damage and accumulation of genetic errors, therefore leading to selective advantage for abnormal cell proliferation¹⁰³. Numerous studies found that CD1 G/A870 single nucleotide polymorphism is associated with two different splice variant transcripts: CD1a and CD1b. CD1a encodes for the full-length native form of the CD1 protein, and CD1b encodes for a truncated alternate CD1 protein¹⁰⁴. This strongly suggests that individuals with numerous copies of the *CCND1-870A* are more likely to bypass the G1-S checkpoint, thus contributing to cancer development¹⁰³. Hiyama et al. evaluated six ORs¹⁰⁵⁻¹¹⁰ from 9 studies¹⁰⁵⁻¹¹³ of the GA genotype vs. the GG genotype which at nucleotide position 870 were < 1, and 7 ORs¹⁰⁵⁻¹¹⁰ for the AA genotype vs GG which were < 1². These results are statistically relevant and suggest that the A allele may be associated with decreased risk for HNC. On the basis of the studies published in 2011, cell cycle regulation may play a role in oral carcinogenesis and the *CCND1* rs9344 polymorphism may be a useful biomarker for oral oncology¹¹⁴. The data reported in the literature indicates that CD1 genotype and protein expression as important risk markers for laryngeal cancer and suggest future trials targeting upstream regulators of CD1 transcription¹⁰⁴.
- Dysfunction in the *P53 tumour suppressor gene* (located at 17p13) is implicated in many cancers, including HNC, and has received the most attention. The production of *p53* is increased in response to cellular insults or DNA damage, and *p53* then induces cell cycle arrest at the G1/S junction. If the damage is irreparable, *p53* can initiate cell death by apoptosis. The steady-state concentration of *p53* in normal cells is low, and the half-life of normal (wild type) *p53* is short. In contrast,

if the *p53* gene is mutated, the genetic product is often present at high concentrations. Even if the concentration is important, mutations in the *TP53* gene frequently occur. Mutations in *p53* are present in 50-60% of head and neck cancers¹¹⁵⁻¹¹⁶. Hiyama published in 2008 that a G-to-C polymorphism in codon 72 of exon 4 results in an Arg-to-Pro substitution. Although both variants are morphologically wild-type, the Pro/Pro genotype is less effective in suppressing cellular transformation¹¹⁷. Individuals with the Pro/Pro genotype showed a higher risk for HNC than individuals with the Arg/Arg genotype in 15¹¹⁸⁻¹³² of 19 studies^{64 118-135}. In 2010, Zhou provided a more precise estimation of the relationship between P53 and head and neck cancers. There was no evidence to suggest that *TP53* codon 72 polymorphisms may be a risk factor for oral carcinoma¹³⁶. However, *TP53* codon 72 polymorphisms may be a risk factor for NPC. The homozygote Pro/Pro genotype could significantly increase susceptibility to NPC, whereas the Arg allele markedly decreases NPC risk¹³⁷. Bradford¹³⁸ and Poeta¹³⁹ published that mutations in tumour protein 53 (*TP53*) may be correlated with aggressive HNSCC disease and relative radioresistance. In the December 2007 issue of the *New England Journal of Medicine*, a team of NIDCR grantees and colleagues evaluated the prognostic value of *TP53* mutations in 420 head-and-neck cancer patients treated with surgery only and whose survival was tracked for several years thereafter. Detecting *TP53* alterations in the tumours of 53% of participants, it was found that these mutations were associated with decreased overall survival.

- *P73* encodes a member of the *p53* family of transcription factors involved in cellular responses to stress and development. It maps to a region on chromosome 1p36 that is frequently deleted in neuroblastoma and other tumours, and thought to contain multiple tumour suppressor genes. The demonstration that this gene is monoallelically expressed (likely from the maternal allele) supports the notion that it is a candidate gene for neuroblastoma. Many transcript variants resulting from alternative splicing and/or use of alternate promoters have been found for this gene, but the biological validity and the full-length nature of some variants have not been determined (RefSeq, Feb 2011). Hypermethylation of *TP73* is reported in nasopharyngeal carcinomas with a frequency of 20%¹⁴⁰. Ferru in 2006¹⁴¹, by comparison to normal thyroid tissue surrounding tumours, observed significant down regulation of *TP73* transcripts in adenomas and in differentiated carcinomas. Chen published in 2004¹⁴² that *p73* expression may be associated with the differentiation of oral stratified squamous epithelium, an early event in human oral carcinogenesis, and associated with the nodal status of patients with oral carcinoma and a possible indicator for malignant change of oral epithelial dysplasia.

Discussion and conclusions

Molecular epidemiologic studies have provided evidence that individual susceptibility to cancer is mediated by both genetic and environmental factors^{4 6-14 33 34 43 50 128 135 140 143}. Interest in the role of genetic polymorphisms in HNC has increased recently, possibly due to advances in DNA analysis technologies or our knowledge of the human genome.

The most intensively-studied genes are those encoding enzymes that metabolize carcinogens and include *GSTM1*, *GSTT1* and *GSTP1*. This is likely because these variants are well characterized, and increased cancer risk associated with these variations is plausible. A considerable amount of work has been done on these genes in relation to risk for HNC. One of the major problems of these studies is that many have a small sample size (< 100 cases or < 100 controls)². Overall, by summarizing the results of the published meta-analyses of the association between genetic polymorphisms and HNC risk, a statistically risk was reported for 3 polymorphisms, namely:

- *Glutathione S-transferase (GSTM1)*, 2 meta-analyses;
- *Glutathione S-transferase (GSTT1)*, 1 meta-analysis;
- *Human microsomal epoxide hydrolase (EPHX1)*, 2 meta-analyses.

As demonstrated by 2 meta-analyses, *GSTM1* deficiency was associated with laryngeal cancer risk. Strikingly, the results showed lack of associations between *GSTM1* null genotypes and laryngeal cancer risk in Caucasians or Asians. The combination of the *GSTM1* null plus the *CYP1A1* (m1m2) variant genotypes increased the risk of oral and pharyngeal cancer²⁶. Previous meta-analysis and pooled analysis have reported an association between the *GSTM1* null genotype and head and neck tumours, but did not analyze ethnic specific or subsite specific differences. Varela-Lema et al.³³ evaluated ethnic specific and subsite specific differences in a pooled analysis, and confirmed that there was no association of the *GSTM1* null genotype with oral and pharyngeal cancers in Caucasians. Although not statistically significant, African American and African populations seemed to be almost twice as likely to have the *GSTM1* null genotype. This lack of statistical significance might also be attributed to the small number of African American and African subjects included in this pooled analysis. There was also an association between *GSTM1* null genotypes and smoking status³⁴.

Deletion of *GSTM1* might contribute to the tumorigenesis and progression of nasopharyngeal cancer²⁶. *GSTT1* deletion might also have an association with increased nasopharyngeal cancer risk, as demonstrated by a meta-analysis. A successive study has demonstrated that the data failed to show a significant association of *GSTT1* null genotype with increased susceptibility to nasopharyngeal cancer. This discrepancy might be due to several reasons. For *GSTT1*, a gene that is highly conserved dur-

ing evolution, major ethnic differences exist in frequency distribution. In East Asia, highest percentages of individuals with the *GSTT1* null genotype have been reported. Interestingly, the incidence of nasopharyngeal cancer is high in East Asia, but is low in other regions worldwide. It thus appears that *GSTT1* deletion may have an association with increased nasopharyngeal cancer risk. Nevertheless, it indicates that although many people in East Asia carry *GSTT1* null genotype, but only a relatively small number of people develop nasopharyngeal cancer, implying that *GSTT1* deletion might not be a key event increasing susceptibility to nasopharyngeal cancer³⁴. Additionally, as only 4 small studies have been published on *GSTT1*, it is likely that the discrepancy may be due to chance because studies with small sample size may have insufficient statistical power to detect a slight effect or may yield a fluctuated risk estimate²⁶. Zhang et al. did not find a significant association between the null genotype of *GSTT1* and oral cancer risk in Asians or Caucasians³⁴. Heterogeneity exists between studies, and a significant multiplicative interaction between the null genotype of *GSTT1* and smoking status has not been found, although small sample sizes may be a reason for the lack of statistical significance.

Two meta-analyses for the *EPHX1* gene demonstrated dissimilar results. The Y113H allele may have a potential protective effect on tobacco-related carcinogenesis of lung and HNC, whereas the homozygous variant of H139R allele, instead, may have a harmful effect⁸³. Moreover, cigarette-smoking status may influence the association of *EPHX1* enzyme activity and the related cancer risk⁸³.

Many primary studies had focalized their attention on alcohol metabolic genes. In particular, Mc Kay et al. found that five genetic variants at three loci, 4q21, 4q23 and 12q24, were significantly associated with HNC risk. The 12q24 variant is positioned in an extended region of LD that contains multiple genes. Candidate genes include the *aldehyde dehydrogenase 2 (ALDH2)*¹⁰.

In accordance with Mc Kay and co-workers, three independent variants of the *ADH* gene have been associated with HNC risk in European populations. The effects of these three variants were generally present for each HNC subsites, but more pronounced in esophageal cancers and males. Strong heterogeneity was found with rs1229984 when stratifying by alcohol consumption. Notably, an association was observed in “ever drinkers-never smokers”, but not in “never drinkers-ever smokers”, suggesting that the effect with the rs1229984 variant is mediated through alcohol rather than tobacco smoking. In contrast, the lack of heterogeneity for rs1573496 when stratifying by alcohol use may imply differences in the mechanism of carcinogenesis among these *ADH* variants¹⁰. Several studies have suggested rs1229984 may influence alcohol consumption behaviour^{10 144-147}.

The integration of genome-based knowledge into healthcare has the potential to improve primary and secondary

prevention¹⁴⁸. Among the greatest promises of genomic medicine is that the unravelling of the genetic origins of common diseases will eventually lead to individualized medicine, in which prevention and treatment strategies are personalized on the basis of the results of predictive genetic tests. Findings from meta-analyses of genetic association studies have the potential to provide a comprehensive view of the impact of genetic risk factors in disease aetiology, especially when exploring gene-environment interactions¹¹.

References

- 1 Szymańska K, Levi JE, Menezes A, et al. *TP53 and EGFR mutations in combination with lifestyle risk factors in tumours of the upper aerodigestive tract from South America*. *Carcinogenesis* 2010;31:1054-9.
- 2 Hiyama T, Yoshihara M, Tanaka S, et al. *Genetic polymorphisms and head and neck cancer risk* (Review). *Int J Oncol* 2008;32:945-73.
- 3 Hashibe M, Brennan P, Benhamou S, et al. *Alcohol drinking in never users of tobacco, cigarette smoking in never drinkers, and the risk of head and neck cancer: pooled analysis in the International Head and Neck Cancer Epidemiology Consortium*. *J Natl Cancer Inst* 2007;16:99:777-89.
- 4 Marron M, Boffetta P, Zhang ZF, et al. *Cessation of alcohol drinking, tobacco smoking and the reversal of head and neck cancer risk*. *Int J Epidemiol* 2010;39:182-96.
- 5 Chuang SC, Jenab M, Heck JE, et al. *Diet and the risk of head and neck cancer: a pooled analysis in the INHANCE consortium*. *Cancer Causes Control* 2012;23:69-88.
- 6 Stott-Miller M, Chen C, Chuang SC, et al. *History of diabetes and risk of head and neck cancer: a pooled analysis from the international head and neck cancer epidemiology consortium*. *Cancer Epidemiol Biomarkers Prev* 2012;21:294-304.
- 7 Nicolotti N, Chuang SC, Cadoni G, et al. *Recreational physical activity and risk of head and neck cancer: a pooled analysis within the international head and neck cancer epidemiology (INHANCE) Consortium*. *Eur J Epidemiol* 2011;26:619-28.
- 8 De Feo E, Rowell J, Cadoni G, et al. *A case-control study on the effect of apolipoprotein E genotype on head and neck cancer risk*. *Cancer Epidemiol Biomarkers Prev* 2010;19:2839-46.
- 9 Negri E, Boffetta P, Berthiller J, et al. *Family history of cancer: pooled analysis in the International Head and Neck Cancer Epidemiology Consortium*. *Int J Cancer* 2009;124:394-401.
- 10 McKay JD, Truong T, Gaborieau V, et al. *A genome-wide association study of upper aerodigestive tract cancers conducted within the INHANCE consortium*. *PLoS Genet* 2011;7:e1001333.
- 11 Gianfagna F, De Feo E, van Duijn CM, et al. *A systematic review of meta-analyses on gene polymorphisms and gastric cancer risk*. *Curr Genomics* 2008;9:361-74.
- 12 Matthias C, Bockmuhl U, Jahnke V, et al. *Polymorphism in cytochrome P450 CYP2D6, CYP1A1, CYP2E1 and glutathione S-transferase, GSTM1, GSTM3, GSTT1 and susceptibility to tobacco-related cancers: studies in upper aerodigestive tract cancers*. *Pharmacogenetics* 1998;8:91-100.
- 13 Katoh T, Kaneko S, Kohshi K, et al. *Genetic polymorphisms of tobacco- and alcohol-related metabolizing enzymes and oral cavity cancer*. *Int J Cancer* 1999;83:606-9.
- 14 Morita S, Yano M, Tsujinaka T, et al. *Genetic polymorphisms of drug-metabolizing enzymes and susceptibility to head-and-neck squamous-cell carcinoma*. *Int J Cancer* 1999;80:685-8.
- 15 Marques CFS, Koifman S, Koifman RJ, et al. *Influence of CYP1A1, CYP2E1, GSTM3, and NAT2 genetic polymorphisms in oral cancer susceptibility: results from a case-control study in Rio de Janeiro*. *Oral Oncol* 2006;42:632-7.
- 16 Sugimura T, Kumimoto H, Tohnai I, et al. *Gene-environment interaction involved in oral carcinogenesis: molecular epidemiological study for metabolic and DNA repair gene polymorphisms*. *J Oral Pathol Med* 2006;35:11-8.
- 17 Gattas GJF, De Carvalho MB, Siraque MS, et al. *Genetic polymorphisms of CYP1A1, CYP2E1, GSTM1, and GSTT1 associated with head and neck cancer*. *Head Neck* 2006;28:819-26.
- 18 Hung HC, Chuang J, Chien YC, et al. *Genetic polymorphisms of CYP2E1, GSTM1, and GSTT1; environmental factors and risk of oral cancer*. *Cancer Epidemiol Biomarkers Prev* 1997;6:901-5.
- 19 Bouchardy C, Hirvonen A, Coutelle C, et al. *Role of alcohol dehydrogenase 3 and cytochrome P-4502E1 genotypes in susceptibility to cancers of the upper aerodigestive tract*. *Int J Cancer* 2000;87:734-40.
- 20 Neuhaus T, Ko YD, Lorenzen K, et al. *Association of cytochrome P450 2E1 polymorphisms and head and neck squamous cell cancer*. *Toxicol Lett* 2004;151:273-82.
- 21 Hildesheim A, Chen CJ, Caporaso NE, et al. *Cytochrome P4502E1 genetic polymorphisms and risk of nasopharyngeal carcinoma: results from a case-control study conducted in Taiwan*. *Cancer Epidemiol Biomarkers Prev* 1995;4:607-10.
- 22 Hildesheim A, Anderson LM, Chen CJ, et al. *CYP2E1 genetic polymorphisms and risk of nasopharyngeal carcinoma in Taiwan*. *J Natl Cancer Inst* 1997;89:1207-12.
- 23 Gonzalez MV, Alvarez V, Pello MF, et al. *Genetic polymorphism of N-acetyltransferase-2, glutathione S-transferase-M1, and cytochromes P450IIE1 and P450IID6 in the susceptibility to head and neck cancer*. *J Clin Pathol* 1998;51:294-8.
- 24 Kongruttanachok N, Sukdikul S, Setavarin S, et al. *Cytochrome P450 2E1 polymorphism and nasopharyngeal carcinoma development in Thailand: a correlative study*. *BMC Cancer* 2001;1:4.
- 25 Gajecka M, Rydzanicz M, Jaskula-Sztul R, et al. *CYP1A1, CYP2D6, CYP2E1, NAT2, GSTM1 and GSTT1 polymorphisms or their combinations are associated with the increased risk of the laryngeal squamous cell carcinoma*. *Mutat Res* 2005;574:112-23.
- 26 Zhuo WL, Wang Y, Zhuo XL, et al. *Polymorphisms of CYP1A1 and GSTM1 and laryngeal cancer risk: evidence-based meta-analyses*. *J Cancer Res Clin Oncol* 2009;135:1081-90.
- 27 Park JY, Muscat JE, Ren Q, et al. *CYP1A1 and GSTM1 polymorphisms and oral cancer risk*. *Cancer Epidemiol Biomarkers Prev* 1997;6:791-7.
- 28 Sato M, Sato T, Izumo T et al. *Genetically high susceptibility to oral squamous cell carcinoma in terms of combined genotyping of CYP1A1 and GSTM1 genes*. *Oral Oncol* 2000;36:267-71.
- 29 Sreelekha TT, Ramadas K, Pandey M, et al. *Genetic polymorphisms of CYP1A1, GSTM1 and GSTT1 genes in Indian oral cancer*. *Oral Oncol* 2001;37:593-8.

- ³⁰ Varzim G, Monteiro E, Silva RA, et al. *CYP1A1 and XRCC1 gene polymorphisms in SCC of the larynx*. *Eur J Cancer Prev* 2003;12:495-9.
- ³¹ Ruwali M, Parmar D. *Association of functionally important polymorphisms in cytochrome P450s with squamous cell carcinoma of head and neck*. *Indian J Exp Biol* 2010;48:651-65.
- ³² Boccia S, Cadoni G, Sayed-Tabatabaei FA, et al. *CYP1A1, CYP2E1, GSTM1, GSTT1, EPHX1 exons 3 and 4, and NAT2 polymorphisms, smoking, consumption of alcohol and fruit and vegetables and risk of head and neck cancer*. *J Cancer Res Clin Oncol* 2008;134:93-100.
- ³³ Varela-Lema L, Taioli E, Ruano-Ravina A, et al. *Meta-analysis and pooled analysis of GSTM1 and CYP1A1 polymorphisms and oral and pharyngeal cancers: a HuGE-GSEC review*. *Genet Med* 2008;10:369-84.
- ³⁴ Zhang ZJ, Hao K, Shi R, et al. *Glutathione S-transferase M1 (GSTM1) and glutathione S-transferase T1 (GSTT1) null polymorphisms, smoking, and their interaction in oral cancer: a HuGE review and meta-analysis*. *Am J Epidemiol* 2011;173:847-57.
- ³⁵ Sato M, Sato T, Izumo T, et al. *Genetic polymorphism of drug-metabolizing enzymes and susceptibility to oral cancer*. *Carcinogenesis* 1999;20:1927-31.
- ³⁶ Trizna Z, Clayman GL, Spitz MR, et al. *Glutathione S-transferase genotypes as risk factors for head and neck cancer*. *Am J Surg* 1995;170:499-501.
- ³⁷ Coutelle C, Ward PJ, Fleury B, et al. *Laryngeal and oropharyngeal cancer, and alcohol dehydrogenase 3 and glutathione S-transferase M1 polymorphisms*. *Hum Genet* 1997;99:319-25.
- ³⁸ Lafuente A, Maristany M, Arias C, et al. *Glutathione and glutathione S-transferase in human squamous cell carcinomas of the larynx and GSTM1 dependent risk*. *Anticancer Res* 1998;18:107-11.
- ³⁹ Cheng L, Sturgis EM, Eicher SA, et al. *Glutathione-S-transferase polymorphisms and risk of squamous-cell carcinoma of the head and neck*. *Int J Cancer* 1999;84:220-4.
- ⁴⁰ Nomura T, Noma H, Shibahara T, et al. *Aldehyde dehydrogenase 2 and glutathione S-transferase M1 polymorphisms in relation to the risk for oral cancer in Japanese drinkers*. *Oral Oncol* 2000;36:42-6.
- ⁴¹ Park JY, Muscat JE, Kaur T, et al. *Comparison of GSTM polymorphisms and risk for oral cancer between African-Americans and Caucasians*. *Pharmacogenetics* 2000;10:123-31.
- ⁴² Hong YJ, Lee JK, Lee GH, et al. *Influence of glutathione S-transferase M1 and T1 genotypes on larynx cancer risk among Korean smokers*. *Clin Chem Lab Med* 2000;38:917-9.
- ⁴³ Kietthubthaw S, Sriplung H, Au WW. *Genetic and environmental interactions on oral cancer in Southern Thailand*. *Environ Mol Mutagen* 2001;37:111-6.
- ⁴⁴ Buch SC, Notani PN, Bhisey RA. *Polymorphism at GSTM1, GSTM3 and GSTT1 gene loci and susceptibility to oral cancer in an Indian population*. *Carcinogenesis* 2002;23:803-7.
- ⁴⁵ Bardakci F, Canbay E, Degerli N, et al. *Relationship of tobacco smoking with GSTM1 gene polymorphism in laryngeal cancer*. *J Cell Mol Med* 2003;7:307-12.
- ⁴⁶ Drummond SN, De Marco L, Noronha JCM, et al. *GSTM1 polymorphism and oral squamous cell carcinoma*. *Oral Oncol* 2004;40:52-5.
- ⁴⁷ Peters ES, McClean MD, Marsit CJ, et al. *Glutathione S-transferase polymorphisms and the synergy of alcohol and tobacco in oral, pharyngeal, and laryngeal carcinoma*. *Cancer Epidemiol Biomarkers Prev* 2006;15:2196-202.
- ⁴⁸ Acar H, Ozturk K, Muslumanoglu MH, et al. *Relation of glutathione S-transferase genotypes (GSTM1 and GSTT1) to laryngeal squamous cell carcinoma risk*. *Cancer Genet Cytogenet* 2006;169:89-93.
- ⁴⁹ Capoluongo E, Almadori G, Concolino P, et al. *GSTT1 and GSTM1 allelic polymorphisms in head and neck cancer patients from Italian Lazio region*. *Clin Chem Acta* 2007;376:174-8.
- ⁵⁰ Hashibe M, Brennan P, Strange RC, et al. *Meta- and pooled analyses of GSTM1, GSTT1, GSTP1, and CYP1A1 genotypes and risk of head and neck cancer*. *Cancer Epidemiol Biomarkers Prev* 2003;12:1509-17.
- ⁵¹ Tripathy CB, Roy N. *Meta-analysis of glutathione S-transferase M1 genotype and risk toward head and neck cancer*. *Head Neck* 2006;28:217-24.
- ⁵² Jahnke V, Matthias C, Fryer A, et al. *Glutathione S-transferase and cytochrome-P450 polymorphism as risk factors for squamous cell carcinoma of the larynx*. *Am J Surg* 1996;172:671-3.
- ⁵³ Sharma A, Mishra A, Das BC, et al. *Genetic polymorphism at GSTM1 and GSTT1 gene loci and susceptibility to oral cancer*. *Neoplasma* 2006;53:309-15.
- ⁵⁴ Hamel N, Karimi S, Hebert-Blouin MN, et al. *Increased risk of head and neck cancer in association with GSTT1 nullizygosity for individuals with low exposure to tobacco*. *Int J Cancer* 2000;87:452-4.
- ⁵⁵ Drummond SN, Gomez RS, Noronha JCM, et al. *Association between GSTT-1 gene deletion and the susceptibility to oral squamous cell carcinoma in cigarette-smoking subjects*. *Oral Oncol* 2005;41:515-9.
- ⁵⁶ Sikdar N, Paul RR, Roy B. *Glutathione S-transferase M3 (A/A) genotype as a risk factor for oral cancer and leukoplakia among Indian tobacco smokers*. *Int J Cancer* 2004;109:95-101.
- ⁵⁷ Matthias C, Bockmuhl U, Jahnke V, et al. *The glutathione S-transferase GSTP1 polymorphism: effects on susceptibility to oral/pharyngeal and laryngeal carcinomas*. *Pharmacogenetics* 1998;8:1-6.
- ⁵⁸ Park JY, Schantz SP, Stern JC, et al. *Association between glutathione S-transferase π genetic polymorphisms and oral cancer risk*. *Pharmacogenetics* 1999;9:497-504.
- ⁵⁹ Katoh T, Kaneko S, Takasawa S, et al. *Human glutathione S-transferase P1 polymorphism and susceptibility to smoking related epithelial cancer; oral, lung, gastric, colorectal and urothelial cancer*. *Pharmacogenetics* 1999;9:165-9.
- ⁶⁰ Olshan AF, Weissler MC, Watson MA, et al. *GSTM1, GSTT1, GSTP1, CYP1A1, and NAT1 polymorphisms, tobacco use, and the risk of head and neck cancer*. *Cancer Epidemiol Biomarkers Prev* 2000;9:185-91.
- ⁶¹ McWilliams JE, Evans AJ, Beer TM, et al. *Genetic polymorphisms in head and neck cancer risk*. *Head Neck* 2000;22:609-17.
- ⁶² Evans AJ, Henner WD, Eilers KM, et al. *Polymorphisms of GSTT1 and related genes in head and neck cancer risk*. *Head Neck* 2004;26:63-70.

- 63 Ko Y, Abel J, Harth V, et al. *Association of CYP1B1 codon 432 mutant allele in head and neck squamous cell cancer is reflected by somatic mutations of p53 in tumor tissue.* *Cancer Res* 2001;61:4398-404.
- 64 Cheng YJ, Chien YC, Hildesheim A, et al. *No association between genetic polymorphisms of CYP1A1, GSTM1, GSTT1, GSTP1, NAT2, and nasopharyngeal carcinoma in Taiwan.* *Cancer Epidemiol Biomarkers Prev* 2003;12:179-80.
- 65 Jourenkova-Mironova N, Voho A, Bouchardy C, et al. *Glutathione S-transferase GSTM1, GSTM3, GSTP1 and GSTT1 genotypes and the risk of smoking-related oral and pharyngeal cancers.* *Int J Cancer* 1999;81:44-8.
- 66 Cabelguenne A, Loriot MA, Stucker I, et al. *Glutathione associated enzymes in head and neck squamous cell carcinoma and response to cisplatin-based neoadjuvant chemotherapy.* *Int J Cancer* 2001;93:725-30.
- 67 To-Figueras J, Gene M, Gomez-Catalan J, et al. *Microsomal epoxide hydrolase and glutathione S-transferase polymorphisms in relation to laryngeal carcinoma risk.* *Cancer Lett* 2001;87:95-101.
- 68 Majumder M, Sikdar N, Paul RR, et al. *Increased risk of oral leukoplakia and cancer among mixed tobacco users carrying XRCC1 variant haplotypes and cancer among smokers carrying two risk genotypes: one of each of two loci, GSTM3 and XRCC1 (codon 280).* *Cancer Epidemiol Biomarkers Prev* 2005;14:2106-12.
- 69 Oude Ophuis M, Manni JJ, Peters WHM. *Glutathione S-transferase T1 null polymorphism and the risk for head and neck cancer.* *Acta Otolaryngol* 2006;126:311-7.
- 70 Jourenkova-Mironova N, Voho A, Bouchardy C, et al. *Glutathione S-transferase GSTM3 and GSTP1 genotypes and larynx cancer risk.* *Cancer Epidemiol Biomarkers Prev* 1999;8:185-8.
- 71 Kelders WPA, Oude Ophuis MB, Roelofs HMJ, et al. *The association between glutathione S-transferase P1 genotype and plasma level in head and neck.* *Laryngoscope* 2002;112:462-6.
- 72 Amador AG, Righi PD, Radpour S, et al. *Polymorphisms of xenobiotic metabolizing genes in oropharyngeal carcinoma.* *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 2002;93:440-5.
- 73 Oude Ophuis MB, Roelofs HNJ, van den Brandt PA, et al. *Polymorphisms of the glutathione S-transferase P1 gene and head and neck cancer susceptibility.* *Head Neck* 2003;25:37-43.
- 74 Cho CG, Lee SK, Nam SY, et al. *Association of the GSTP1 and NQO1 polymorphisms and head and neck squamous cell carcinoma risk.* *J Korean Med Sci* 2006;21:1075-9.
- 75 Agúndez JA. *Polymorphisms of human N-acetyltransferases and cancer risk.* *Curr Drug Metab* 2008;9:520-31.
- 76 Hahn M, Hagedorn G, Kuhlisch E, et al. *Genetic polymorphisms of drug-metabolizing enzymes and susceptibility to oral cavity cancer.* *Oral Oncol* 2002;38:486-90.
- 77 Katoh T, Kaneko S, Boissy R, et al. *A pilot study testing the association between N-acetyltransferases 1 and 2 and risk of oral squamous cell carcinoma in Japanese people.* *Carcinogenesis* 1998;19:1803-7.
- 78 Varzim G, Monteiro E, Siva R, et al. *Polymorphisms of arylamine N-acetyltransferase (NAT1 and NAT2) and larynx cancer susceptibility.* *ORL* 2002;64:206-12.
- 79 Chen C, Ricks S, Doody DR, et al. *N-acetyltransferase 2 polymorphisms, cigarette smoking and alcohol consumption, and oral squamous cell cancer risk.* *Carcinogenesis* 2001;22:1993-19.
- 80 Ying XJ, Dong P, Shen B, et al. *Possible association of NAT2 polymorphism with laryngeal cancer risk: an evidence-based meta-analysis.* *J Cancer Res Clin Oncol* 2011;137:1661-7.
- 81 Buch SC, Nazar-Stewart V, Weissfeld JL, et al. *Case-control study of oral and oropharyngeal cancer in whites and genetic variation in eight metabolic enzymes.* *Head Neck* 2008;30:1139-47.
- 82 Demokan S, Suoglu Y, Gözeler M, et al. *N-acetyltransferase 1 and 2 gene sequence variants and risk of head and neck cancer.* *Mol Biol Rep* 2010;37:3217-26.
- 83 Li X, Hu Z, Qu X, et al. *Putative EPHX1 enzyme activity is related with risk of lung and upper aerodigestive tract cancers: a comprehensive meta-analysis.* *PLoS One* 2011;6:e14749.
- 84 Yokoyama A, Muramatsu T, Ohmori T, et al. *Alcohol-related cancers and aldehyde dehydrogenase-2 in Japanese alcoholics.* *Carcinogenesis* 1998;19:1383-7.
- 85 Yokoyama A, Muramatsu T, Ohmori T, et al. *Alcohol and aldehyde dehydrogenase gene polymorphisms and oropharyngeal, esophageal and stomach cancers in Japanese alcoholics.* *Carcinogenesis* 2001;22:433-9.
- 86 Asakage T, Yokoyama A, Haneda T, et al. *Genetic polymorphisms of alcohol and aldehyde dehydrogenases, and drinking, smoking and diet in Japanese men with oral and pharyngeal squamous cell carcinoma.* *Carcinogenesis* 2007;28:865-74.
- 87 Tiemersma EW, Wark PA, Ocke MC, et al. *Alcohol consumption, alcohol dehydrogenase 3 polymorphism, and colorectal adenomas.* *Cancer Epidemiol Biomarkers Prev* 2003;12:419-25.
- 88 Harty LC, Caporaso NE, Hayes RB, et al. *Alcohol dehydrogenase 3 genotype and risk of oral cavity and pharyngeal cancers.* *J Natl Cancer Inst* 1997;89:1968-75.
- 89 Olshan AF, Weissler MC, Watson MA, et al. *Risk of head and neck cancer and the alcohol dehydrogenase 3 genotype.* *Carcinogenesis* 2001;22:57-61.
- 90 Sturgis EM, Dahlstrom KR, Guan Y, et al. *Alcohol dehydrogenase 3 genotype is not associated with risk of squamous cell carcinoma of the oral cavity and pharynx.* *Cancer Epidemiol Biomarkers Prev* 2001;10:273-5.
- 91 Schwartz SM, Doody DR, Fitzgibbons ED, et al. *Oral squamous cell cancer risk in relation to alcohol consumption and alcohol dehydrogenase-3 genotypes.* *Cancer Epidemiol Biomarkers Prev* 2001;10:1137-44.
- 92 Zavras AI, Wu T, Laskaris G, et al. *Interaction between a single nucleotide polymorphism in the alcohol dehydrogenase 3 gene, alcohol consumption and oral cancer risk.* *Int J Cancer* 2002;97:526-30.
- 93 Nishimoto IN, Pinheiro NA, Rogatto SR, et al. *Alcohol dehydrogenase 3 genotype as a risk factor for upper aerodigestive tract cancers.* *Arch Otolaryngol Head Neck Surg* 2004;130:78-82.
- 94 González CA, Sala N, Capellá G. *Genetic susceptibility and gastric cancer risk.* *Int J Cancer* 2002;100:249-60.
- 95 Drapkin R, Sancar A, Reinberg D. *Where transcription meets repair.* *Cell* 1994;77:9-12.
- 96 Qiao Y, Spitz MR, Shen H, et al. *Modulation of repair of ultraviolet damage in the host-cell reactivation assay by poly-*

- morphic XPC and XPD/ERCC2 genotypes*. *Carcinogenesis* 2002;23:295-9.
- ⁹⁷ Kietthubthew S, Sriplung H, Au WW, et al. *Polymorphism in DNA repair genes and oral squamous cell carcinoma in Thailand*. *Int J Hyg Environ Health* 2006;209:21-9.
- ⁹⁸ Majumder M, Sikdar N, Ghosh S, et al. *Polymorphisms at XPD and XRCC1 DNA repair loci and increased risk of oral leukoplakia and cancer among NAT2 slow acetylators*. *Int J Cancer* 2007;120:2148-56.
- ⁹⁹ Gajecka M, Rydzanicz M, Jaskula-Sztul R, et al. *Reduced DNA repair capacity in laryngeal cancer subjects*. *Adv Otorhinolaryngol* 2005;62:25-37.
- ¹⁰⁰ Sturgis EM, Zheng R, Li L, et al. *XPD/ERCC2 polymorphisms and risk of head and neck cancer: a case-control analysis*. *Carcinogenesis* 2000;21:2219-23.
- ¹⁰¹ Ramachandran S, Ramadas K, Hariharan R, et al. *Single nucleotide polymorphisms of DNA repair genes XRCC1 and XPD and its molecular mapping in Indian oral cancer*. *Oral Oncol* 2006;42:350-62.
- ¹⁰² Huang WY, Olshan AF, Schwartz SM, et al. *Selected genetic polymorphisms in MGMT, XRCC1, XPD, and XRCC3 and risk of head and neck cancer: a pooled analysis*. *Cancer Epidemiol Biomarkers Prev* 2005;14:1747-53.
- ¹⁰³ Pabalan N, Bapat B, Sung L, et al. *Cyclin D1 Pro241Pro (CCND1-G870A) polymorphism is associated with increased cancer risk in human populations: a meta-analysis*. *Cancer Epidemiol Biomarkers Prev* 2008;17:2773-81.
- ¹⁰⁴ Papadimitrakopoulou V, Izzo JG, Liu DD, et al. *Cyclin D1 and cancer development in laryngeal premalignancy patients*. *Cancer Prev Res (Phila)* 2009;2:14-21.
- ¹⁰⁵ Deng L, Zhao XR, Pan KF, et al. *Cyclin D1 polymorphism and susceptibility to NPC using DHPLC*. *Acta Biochem Biophys Sinica* 2002;34:16-20.
- ¹⁰⁶ Wong YK, Lin SC, Chang CS, et al. *Cyclin D1 genotype in areca-associated oral squamous cell carcinoma*. *J Oral Pathol Med* 2003;32:265-70.
- ¹⁰⁷ Nishimoto IN, Pinheiro NA, Rogatto SR, et al. *Cyclin D1 gene polymorphism as a risk factor for squamous cell carcinoma of the upper aerodigestive system in non-alcoholics*. *Oral Oncol* 2004;40:604-10.
- ¹⁰⁸ Monteiro E, Varzim G, Pires AM, et al. *Cyclin D1 A870G polymorphism and amplification in laryngeal squamous cell carcinoma: implications of tumor localization and tobacco exposure*. *Cancer Detect Prev* 2004;28:237-43.
- ¹⁰⁹ Holley SL, Matthias C, Jahnke V, et al. *Association of cyclin D1 polymorphism with increased susceptibility to oral squamous cell carcinoma*. *Oral Oncol* 2005;41:156-60.
- ¹¹⁰ Catarino RJ, Breda E, Coelho V, et al. *Association of the A870G cyclin D1 gene polymorphism with genetic susceptibility to nasopharyngeal carcinoma*. *Head Neck* 2006;28:603-8.
- ¹¹¹ Zheng Y, Shen H, Sturgis EM, et al. *Cyclin D1 polymorphism and risk for squamous cell carcinoma of the head and neck: a case-control study*. *Carcinogenesis* 2001;22:1195-9.
- ¹¹² Rydzanicz M, Golusinski P, Mielcarek-Kuchta D, et al. *Cyclin-D1 gene (CCND1) polymorphism and the risk of squamous cell carcinoma of the larynx*. *Eur Arch Otorhinolaryngol* 2006;263:43-8.
- ¹¹³ Sathyan KM, Nalinakumari KR, Abraham T, et al. *Influence of single nucleotide polymorphisms in H-Ras and cyclin D1 genes on oral cancer susceptibility*. *Oral Oncol* 2006;42:607-13.
- ¹¹⁴ Tsai MH, Tsai CW, Tsou YA, et al. *Significant association of cyclin D1 single nucleotide polymorphisms with oral cancer in taiwan*. *Anticancer Res* 2011;31:227-31.
- ¹¹⁵ Golusinski P, Lamperska K, Pazdrowski J, et al. *Analysis of mutations within the TP53 gene in patients with squamous cell carcinoma of the head and neck*. *Otolaryngol Pol* 2011;65:114-21.
- ¹¹⁶ Galli P, Cadoni G, Volante M, et al. *A case-control study on the combined effects of p53 and p73 polymorphisms on head and neck cancer risk in an Italian population*. *BMC Cancer* 2009;9:137.
- ¹¹⁷ Thomas M, Kalita A, Labrecque S, et al. *Two polymorphic variants of wild-type p53 differ biochemically and biologically*. *Mol Cell Biol* 1999;19:1092-1100.
- ¹¹⁸ Lu J, Wang LE, Xiong P, et al. *172G > T variant in the 5' untranslated region of DNA repair gene RAD51 reduces risk of squamous cell carcinoma of the head and neck and interacts with a P53 codon 72 variant*. *Carcinogenesis* 2007;28:988-94.
- ¹¹⁹ Birgander R, Sjalander A, Zhou Z, et al. *p53 polymorphisms and haplotypes in nasopharyngeal cancer*. *Hum Hered* 1996;46:49-54.
- ¹²⁰ Golovleva I, Birgander R, Sjalander A, et al. *Interferon-α and P53 alleles involved in nasopharyngeal carcinoma*. *Carcinogenesis* 1997;18:645-9.
- ¹²¹ Hamel N, Black MJ, Ghadirian P, et al. *No association between p53 codon 72 polymorphism and risk of squamous cell carcinoma of the head and neck*. *Br J Cancer* 2000;82:757-9.
- ¹²² Summersgill KF, Smith EM, Kirchner HL, et al. *p53 polymorphism, human papillomavirus infection in the oral cavity, and oral cancer*. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 2000;90:334-9.
- ¹²³ Tandle AT, Sanghvi V, Saranath D. *Determination of p53 genotypes in oral cancer patients from India*. *Br J Cancer* 2001;84:739-42.
- ¹²⁴ Shen H, Zheng Y, Sturgis EM, et al. *P53 codon 72 polymorphism and risk of squamous cell carcinoma of the head and neck: a case-control study*. *Cancer Lett* 2002;183:123-30.
- ¹²⁵ Drummond SN, De Marco L, Pordeus IDA, et al. *TP53 codon 72 polymorphism in oral squamous cell carcinoma*. *Anticancer Res* 2002;22:3379-81.
- ¹²⁶ Tiwawech D, Srivantanakul P, Karaluk A, et al. *The p53 codon 72 polymorphism in Thai nasopharyngeal carcinoma*. *Cancer Lett* 2003;198:69-75.
- ¹²⁷ Kietthubthew S, Sriplung H, Au WW, et al. *The p53 codon 72 polymorphism and risk of oral cancer in Southern Thailand*. *Asian Pac J Cancer Prev* 2003;4:209-14.
- ¹²⁸ Katiyar S, Thelma BK, Murthy NS, et al. *Polymorphism of the p53 codon 72 arg/pro and the risk of HPV type 16/18-associated cervical and oral cancer in India*. *Mol Cell Biochem* 2003;252:117-24.
- ¹²⁹ Cortezzi SS, Provazzi PJ, Sobrinho JS, et al. *Analysis of human papillomavirus prevalence and TP53 polymorphism in head and neck squamous cell carcinomas*. *Cancer Genet Cytogenet* 2004;150:44-9.
- ¹³⁰ Twu CW, Jiang RS, Shu CH, et al. *Association of p53 codon 72 polymorphism with risk of hypopharyngeal sq-*

- amous cell carcinoma in Taiwan. *J Formos Med Assoc* 2006;105:99-104.
- ¹³¹ Sousa H, Santos AM, Catarino R, et al. *Linkage of TP53 codon 72 pro/pro genotype as predictive factor for nasopharyngeal carcinoma development*. *Eur J Cancer Prev* 2006;15:362-6.
- ¹³² Yung WCW, Ng MH, Sham JST, et al. *p53 codon 72 polymorphism in nasopharyngeal carcinoma*. *Cancer Genet Cytogenet* 1997;93:181-2.
- ¹³³ Sourvinos G, Rizos E, Spandidos DA. *p53 codon 72 polymorphism is linked to the development and not the progression of benign and malignant laryngeal tumours*. *Oral Oncol* 2001;37:572-8.
- ¹³⁴ Scheckenback K, Lieven O, Gotte K, et al. *p53 codon 72 polymorphic variants, loss of allele-specific transcription, and human papilloma virus 16 and/or 18 E6 messenger RNA expression in squamous cell carcinomas of the head and neck*. *Cancer Epidemiol Biomarkers Prev* 2004;13:1805-9.
- ¹³⁵ Perrone F, Mariani L, Pastore E, et al. *p53 codon 72 polymorphisms in human papillomavirus-negative and papillomavirus-positive squamous cell carcinomas of the oropharynx*. *Cancer* 2007;109:2461-5.
- ¹³⁶ Zhuo XL, Li Q, Zhou Y, et al. *Study on TP53 codon 72 polymorphisms with oral carcinoma susceptibility*. *Arch Med Res* 2009;40:625-34.
- ¹³⁷ Zhuo XL, Cai L, Xiang ZL, et al. *TP53 codon 72 polymorphism contributes to nasopharyngeal cancer susceptibility: a meta-analysis*. *Arch Med Res* 2009;40:299-305.
- ¹³⁸ Bradford CR, Zhu S, Ogawa H, et al. *P53 mutation correlates with cisplatin sensitivity in head and neck squamous cell carcinoma lines*. *Head Neck* 2003;25:654-61.
- ¹³⁹ Poeta ML, Manola J, Goldwasser MA, et al. *TP53 mutations and survival in squamous-cell carcinoma of the head and neck*. *N Engl J Med* 2007;357:2552-61.
- ¹⁴⁰ Worsham MJ, Chen KM, Meduri V, et al. *Epigenetic events of disease progression in head and neck squamous cell carcinoma*. *Arch Otolaryngol Head Neck Surg* 2006;132:668-77.
- ¹⁴¹ Ferru A, Denis S, Guilhot J, et al. *Expression of TAp73 and DeltaNp73 isoform transcripts in thyroid tumours*. *Eur J Surg Oncol* 2006;32:228-30.
- ¹⁴² Chen YK, Hsue SS, Lin LM. *p73 expression for human buccal epithelial dysplasia and squamous cell carcinoma: does it correlate with nodal status of carcinoma and is there a relationship with malignant change of epithelial dysplasia?* *Head Neck* 2004;26:945-52.
- ¹⁴³ Boccia S, Cadoni G, La Torre G, et al. *A case-control study investigating the role of sulfotransferase 1A1 polymorphism in head and neck cancer*. *J Cancer Res Clin Oncol* 2006;132:466-72.
- ¹⁴⁴ Macgregor S, Lind PA, Bucholz KK, et al. *Associations of ADH and ALDH2 gene variation with self report alcohol reactions consumption and dependence: an integrated analysis*. *Hum Mol Genet* 2009;18:580-93.
- ¹⁴⁵ Tolstrup JS, Nordestgaard BG, Rasmussen S, et al. *Alcoholism and alcohol drinking habits predicted from alcohol dehydrogenase genes*. *Pharmacogenomics J* 2008;8:220-7.
- ¹⁴⁶ Zuccolo L, Fitz-Simon N, Gray R, et al. *A non-synonymous variant in ADH1B is strongly associated with prenatal alcohol use in a European sample of pregnant women*. *Hum Mol Genet* 2009;15:4457-66.
- ¹⁴⁷ Luo X, Kranzler HR, Zuo L, et al. *Diplotype trend regression analysis of the ADH gene cluster and the ALDH2 gene: multiple significant associations with alcohol dependence*. *Am J Hum Genet* 2006;78:973-87.
- ¹⁴⁸ Ricciardi W, Boccia S. *Assessment of genomics as a priority for Public Health*. *Eurohealth* 2007;13:7-9.
- ¹⁴⁹ Hu Z, Ma H, Chen F, et al. *XRCC1 polymorphisms and cancer risk: a meta-analysis of 38 case-control studies*. *Cancer Epidemiol Biomarkers Prev* 2005;14:1810-8.

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