RESEARCH

MIB-1, BcI-2 and p53 in odontogenic myxomas of the jaws

Mib-1, Bcl-2 e p53 nei mixomi odontogeni dei mascellari

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SUMMARY

Odontogenic myxoma is a rare benign neoplasm occurring in the jaws. Microscopically, it is composed of spindle or stellate-shaped cells arranged in a mucinous matrix. In some cases (20%), odontogenic epithelial islands may be found. The Authors evaluated p53, MIB-1, and Bcl-2 expressed by the epithelial and stromal elements in 12 cases of odontogenic myxoma of the jaws. The cells of the odontogenic epithelium were positive for Bcl-2, p53 and MIB-1. The stromal cell showed a high positivity for MIB-1. Proliferation of both the epithelial and stromal components could be related to the growth of this odontogenic tumour

KEY WORDS: Jaws • Benign neoplasm • Odontogenic myxoma • Apoptosis • Bcl-2 • MIB-1 • p53

RIASSUNTO

Il mixoma odontogeno (OM) è una rara forma di neoplasia benigna dei mascellari. Al microscopio risulta formata da cellule di forma fusata o stellata disposte in una matrice mucoide. Nel 20% dei casi possono esserci delle isole epiteliali odontogene. In questo lavoro è stata valutata l'espressione di p53, MIB-1, e Bcl-2 nell'epitelio e nello stroma di 12 casi di OM della mascella. Le cellule dell'epitelio sono risultate positive per Bcl-2, p53 e MIB-1, mentre quelle stromali mostrano un'alta positività solo per MIB-1. La proliferazione delle componenti stromali ed epiteliali può essere correlata alla crescita del tumore odontogeno.

PAROLE CHIAVE: Mascellari • Tumori benigni • Mixoma odontogeno • Apoptosi • Bcl-2 • MIB-1 • p53

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Introduction

Odontogenic myxoma (OM) is a relatively rare, benign neoplasm occurring in the jaws 1. OM represents 3-20% of all odontogenic tumours (OT) and, in most studies, OM is the third most frequent OT 1-9. An annual incidence of 0.07 per million has been reported 6. It represents from 1.1% to 3.7% of total surgical specimens submitted to an oral pathology department 1. The incidence of OM is about 1/6 that of ameloblastoma 10. Slootweg and Wittkampf 11 found that OM constituted 0.1% of all specimens studied in a 40-year period. OM is a locally invasive neoplasm that shows little encapsulation and often extends through the bone 12. A high recurrence rate has been reported and local infiltration is thought to account for the high recurrence rate 12. The histogenesis of OM is poorly understood, it is still a matter of debate 13 and the underlying molecular mechanisms of OM remain unknown 14. According to most investigators, it represents a tumour unique to the jaws arising from the mesenchymal part of the tooth germ 1 10. Mutations of the Gs alpha gene are rarely, if ever, associated with sporadic jaw myxoma tumourigenesis 14, also a mutation in the protein kinase A regulatory subunit type 1A (PRKAR1A) has been reported 15. OM may originate from either mesenchymal

elements of the dental papilla, dental follicle, periodontal membrane, or non-dental mesenchyme 1 10. Also myxomas located in the soft tissues of the head and neck have been reported 16. The odontogenic nature of the tumour is suggested by the location within alveolar bone in tooth-bearing regions, the association with missing, incompletely formed or non-erupted teeth, the early age of the patients, the presence of islands and strands of odontogenic epithelium, and clinical behaviour resembling that of ameloblastoma ^{1 10 17}. Several mechanisms may be used by neoplastic cells to provide a growth advantage over normal tissue ¹². Neoplastic cells may present an increased rate of cell division and/or a decreased rate of apoptosis 12. Apoptosis allows control of the number of cells in a given tissue 12. Tumour cells that inhibit apoptosis show a growth advantage over normal tissue and apoptosis is regulated, to a large extent, by the Bcl-2 gene family 12. One hypothesis has been recently put forward that OM may present an alteration in the apoptotic mechanisms that helps the growth of these tumours, and, in fact, an increase in cells staining positively for Bcl-2 was found 12. The production, then, of anti-apoptotic proteins with a decrease in the programmed cell death of the cells constituting OM, may provide a growth advantage in this type of tumour ¹².

Ki-67 antigen expression has been observed in the nuclei of proliferating cells and it can be a marker to estimate the state of tissue growth. It has been reported that Ki-67 antigen expression increases in pre-neoplastic and neoplastic lesions of the oral mucosa, and in all states of high cell turnover ¹⁸ ¹⁹. MIB-1 is the monoclonal antibody that reacts with the epitope of the Ki-67 nuclear antigen in formalin-fixed, paraffin-embedded sections ¹⁸ ¹⁹.

Bcl-2 is an anti-apoptotic protein that seems to be important in cancer development and progression ²⁰. Also p53 seems to be important in oncogenesis ²¹. Ki-67, Bcl-2 and p53 have been extensively studied in neck cancerogenesis, in some types of OTs and salivary gland tumours, in odontogenic cysts, and actinic cheilitis ²¹⁻⁵³. The presence of Bcl-2 seems to be associated with a better prognosis in some tumours, but not in others ⁵⁴⁻⁵⁶.

A correlation between these different factors has been hypothesized. Studies in human breast cancer and in cancer cell lines have shown that p53 can down-regulate Bcl-2 expression ⁵⁷ and that apoptosis induced by p53 can be blocked by Bcl-2, in cultured cancer cells 57. p53 has been shown to down-regulate Bcl-2 via binding to a negative regulatory element outside the Bcl-2 gene promoter⁵⁸. Ravi et al. 40 reported an increased Bcl-2 expression in oral dysplasia and carcinoma. Cruz et al. 43 demonstrated that p53 supra-basal expression was significantly associated with the development of carcinoma. Murti et al. 39 found that the over-expression of p53 protein was significantly more common in severe than in mild epithelial dysplasia, and that p53 expression peaked close to the time of transition from pre-cancer to cancer rather than earlier in the natural history of oral pre-cancer. Apoptosis can be important in tumour growth and prognosis: Langlois et al. 59, in colorectal carcinoma, found that tumours with higher apoptotic counts seemed to have good prognosis and this may mean that neoplasms with higher levels of apoptosis are slower growing. In renal cell carcinoma, Hindermann et al. 60 found a decrease in cells undergoing apoptosis in less differentiated tumours with an increase in the number of tumour cells and of tumour growth. This decrease in apoptosis was correlated with an increase in the proliferative activity. The presence of Ki-67 closely coincided with p53 protein 41. Aim of the present study was to analyse MIB-1, p53 and Bcl-2 in stromal and epithelial cells of OM of the jaws.

Materials and methods

The slides of 12 OM of the jaws were retrieved from the archives of the Institute of Pathology, Polytechnic University of the Marche, Ancona, Italy. The patients comprised 7 females and 5 males, mean age 34.4 years (range 18-47). Overall, 11 lesions were located in the posterior mandible, while one was located in the anterior maxilla; the radiographic appearance was of a multi-locular lesion in 8 cases and a uni-locular lesion in 4. All patients were asymptomatic and the lesion had been discovered on X-rays performed for other causes. All lesions had been treated with wide resections. Two recurrences were reported after 4 and 6 years, respectively. All slides were carefully reviewed and the diagnosis of OM of the jaws was confirmed.

Immuno-histochemical staining for Bcl-2 protein was performed using the following antigen retrieval system. Sec-

tions were deparaffinized in two changes of xylene for 10 minutes each and were then rehydrated through graded alcohols and immersed in 0.3% hydrogen peroxide in methanol for 30 minutes to block endogenous peroxidase activity. Sections were then washed in phosphate-buffered saline (PBS). The tissue sections were placed in a microwave oven (Cooktyronic M720, 700 W, Philips, Andover, MA, USA) in a plastic Coplin jar filled with 10 mM sodium citrate buffer (pH 6.0), at 5-minute intervals, for a total of 10 minutes. At each 5-minute interval, the Coplin jar was taken out of the microwave oven and allowed to cool. Slides were incubated overnight with a 1:60 dilution of the primary mouse anti-human Bcl-2 monoclonal antibody (Dako 124, Glostrup, Denmark). A biotin streptavidin detection system was used with diaminobenzidine as the chromogen. Slides were washed twice with PBS and incubated with the linking reagent (biotinylated anti-immunoglobulins) for 15 minutes, at room temperature. After rinsing in PBS, the slides were incubated with the peroxidase-conjugated streptavidin label for 15 minutes, at room temperature. The sections were again rinsed in PBS and incubated with diaminobenzidine for 10 minutes, in the dark. After chromogen development, slides were washed in two changes of water and counterstained with a 1:10 dilution of haematoxylin. The sections were then dehydrated, cleared in xylene, and mounted. A negative control was performed in all cases by omitting the primary antibody, which, in all instances, resulted in negative immuno-reactivity. Sections from a lymph node with follicular lymphoma were used as positive controls. Normal lymphocytes infiltrating the connective tissue of the cyst wall represented an internal positive control for Bcl-2 immuno-staining. In all positive cases, immuno-reactivity was restricted to the cytoplasm. Immuno-histochemical staining for p53 protein was performed using the following antigen retrieval system. Sections were deparaffinized in two changes of xylene for 10 minutes each and then were rehydrated through graded alcohols and immersed in 0.3% hydrogen peroxide in methanol for 30 minutes to block endogenous peroxidase activity. Sections were then washed in PBS. The tissue sections were placed in a microwave oven (Philips, Cooktyronic M720, 700 W) in a plastic Coplin jar filled with 10 mM sodium citrate buffer (pH 6.0), at 5minute intervals, for a total of 10 minutes. At each 5-minute interval, the fluid level in the Coplin jar was removed from the microwave oven and allowed to cool. Slides were incubated overnight with a 1:50 dilution of the primary mouse anti-human p53 monoclonal antibody (Dako DO-7, Glostrup, Denmark). A biotin-streptavidin detection system was used with diaminobenzidine as the chromogen. Slides were washed twice with PBS and incubated with the linking reagent (biotinylated anti-immunoglobulins) for 15 minutes, at room temperature. After rinsing in PBS, the slides were incubated with the peroxidase-conjugated streptavidin label for 15 minutes, at room temperature. The sections were again rinsed in PBS and incubated with diaminobenzidine for 10 minutes, in the dark. After chromogen development, slides were washed in two changes of water and counterstained with a 1:10 dilution of haematoxylin. The sections were then dehydrated, cleared in xylene, and mounted. A negative control was performed in all cases by omitting the primary antibody, which, in all instances, resulted in negative immunoreactivity. p53 expression and location were evaluated on histological sections using a Leitz Orthoplan

microscope equipped with a X 63 objective and with an eyepiece graticule. Only nuclear staining of epithelial cells was observed and the nuclei with a clear brown colour, regardless of staining intensity, were regarded as p53 positive. For MIB-1 immunostaining the slides were pretreated with 3-aminopropyltriethoxysilane (APES-Sigma, St. Louis, MO, USA), which avoided separation of the section from the slide during incubation in the microwave oven. For each case, a 5 μ m section was cut and placed on a pretreated slide. The staining protocol of this slide consisted in the applications of a series of reagents in the following manner: overnight drying at 37 °C; dewaxing and rehydration; immersion in a plastic box containing 0.01 M citrate buffer at pH 6.0; incubation for 5 min in microwave oven: initially at 750 watts until boiling began, then at 350 watts for the remaining time; incubation for 5 min in microwave oven at 350 watts; cooling for 20 min, at room temperature; washing in running water and, then, in distilled water for 5 min; washing in tris buffer saline (TBS) for 5 min; removal of any excess TBS; addition of primary monoclonal mouse anti-ki-67 antibody (Immunotech, Praha, Czech Republic) diluted 1:25 in TBS; overnight incubation at 4 °C in a humidified room; washing in TBS for 5 min (3 times); addition of secondary prediluted biotinylated anti-mouse antibody (LSAB-Dako) and incubation for 10 min, at room temperature; washing in TBS for 5 min (3 times); addition of prediluted streptavidin-peroxidase complex (LSAB-Dako) and incubation for 10 min, at room temperature; washing in TBS for 5 min (3 times); immersion in 0.05% DAB and 0.01% H₂O₂ in TBS for 2-3 min at room temperature; washing in running water, then in distilled water for 5 min; counterstaining with ethyl-green for 30 min; washing in distilled water for 30 sec; washing in buthanol I for 5 sec; washing in buthanol II for 30 sec; dehydration and mounting in Permount (Biomeda, Foster City, CA, USA)

Results

Macroscopically, the lesions had a myxoid consistency, and were partially surrounded by a thin fibrous capsule. Microscopically, the tumours consisted of spindle and stellate cells dispersed in a faintly basophilic ground substance. The degree of cellularity varied in the central and in the peripheral areas. Strands of apparently inactive odontogenic epithelium, surrounded by collagenous fibres, were present in 3 cases. No mitoses were present. In some cells, nuclear atypia was present. The immunoreactivity for p53 protein was

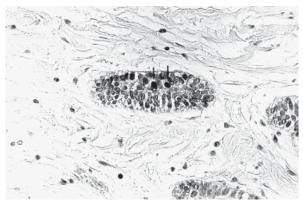


Fig. 1. p53. Positivity of some epithelial cells (arrows) x 200.

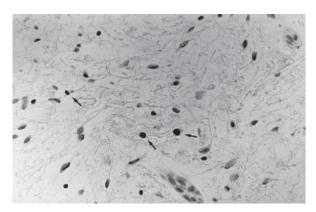


Fig. 2. MIB-1. Positivity of some stromal cells (arrows) x 200.

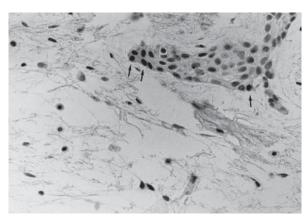


Fig. 3. MIB-1. Positivity of an epithelial cell (arrow) x 200.

present in only 5-10% of the epithelial cells (Fig. 1), while the stromal component was completely negative. The positivity for MIB-1 was overall less than that observed for p53. A higher positivity for MIB-1 of the stromal cells (4%) (Fig. 2) than epithelial cells (1%) (Fig. 3) was observed. With Bcl-2, a strong positivity of the lymphocytes was present: the stroma was completely negative, while only about 2-5% of the epithelial cells were weakly positive (Table I).

Table I. Proliferative activity and oncoprotein expression in odontogenic myxomas.

	Epithelial cells %	Stromal cells %
p53	5-10	0
MIB-1	1	4
Bcl-2	2-5	0

Discussion

OM is an uncommon, benign neoplasm that is thought to be derived from ectomesenchyme and, histologically, is similar to the dental papilla of the developing tooth ¹⁶¹. Due to the fact that most OM have been reported in the tooth-bearing areas of the jaws, there is strong support that this is an ectomesenchymal OT ¹⁶¹. Ultra-structural, biochemical and histochemical studies suggest that the neoplastic spindle cells are fibroblast-like cells called "myxoblasts" with a capability to synthesize and produce a large quantity of mucopolysaccharides ⁶¹. The neoplastic cells show positivity

for vimentin and, in some areas, for muscle specific actin, but, on the contrary, are negative for keratin, desmin, neurospecific enolase, neurofilament and glial fibrillary acidic protein ⁶¹. The majority of the patients have no clinical signs or symptoms ⁶. Pain, dysesthesia, mucosal ulceration, invasion of the soft tissues, tooth mobility may be present ⁶. The epithelium that may be found within the lesion shows positivity to cytokeratin ¹⁹ and this fact is consistent with an odontogenic origin ⁶².

OM is found most frequently in young patients, with a peak incidence in the 2nd and 3rd decades 16. The mandible is more often involved with the ramus and body being equally affected 12. It is a slow growing tumour that can cause marked facial asymmetry 1 61. The tumour may produce symptoms, depending on the site, from tooth mobility to sinus obstruction if the maxilla is involved 161. A very rare malignant variant (myxosarcoma) has been reported 61-70. Macroscopically, OM appears as a soft, lobulated non-encapsulated mass that, on sectioning, has a graying-yellow, glistening, firm, mucoid surface 161. The cut surface fails to bulge beyond the surrounding tissues upon sectioning 161. OM may appear also grossly as mucoid, slimy specimens with a semi-gelatinous consistency 161. In this instance, a capsule is not identified and areas of bone are often present within the cut surface 161. Microscopically, there are stellate or spindle-shaped cells of mesenchymal origin (vimentin positive), loosely arranged in a soft, mucoid matrix, composed of glycosaminoglycans, primarily hyaluronic acid and chondroitin sulfate 12. Hyperchromatic nuclei and mitotic figures are rare, and in about 20% of the lesions, scattered islands and strands of inactive-looking odontogenic epithelium may be found 1 61. These epithelial rests are

mission electron microscopy with the rests of Malassez found in the periodontal ligament surrounding teeth 1 61. The odontogenic epithelium may represent nothing more than a foetal remnant, or it may play an important role in the pathogenesis of OM: it has been suggested that myxoma cells develop as a result of an inductive effect of this odontogenic epithelium 63-69. The strands of odontogenic epithelium, when present, are generally termed "inactive". The present results show, on the contrary, that epithelial cells are positive for Bcl-2, p53 and MIB-1: it is possible to document then the presence of proteins involved in the cell replication or the positivity of a proliferation index. From these data, it can be hypothesized that the growth of the tumour could be related also to this epithelial component. Perhaps it is worthwhile pointing out the fact that the cases where the odontogenic epithelium was present had a larger diameter than the others. The stromal cells showed a higher positivity for MIB-1, and this fact could confirm the results of previous studies that showed that OM mainly grows due to the activity of the stromal cells that have been found to be actively proliferating 64-69. In fact, fine structural features indicate that the connective tissue cells of myxoma are active cells synthesizing and excreting an extracellular matrix 64 68 69. OM could also present dysregulated apoptotic mechanisms that assist the neoplastic growth 12. Both proliferation and matrix production could then determine an increase in tumour size 65.

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References

Buchner A, Odell EW. Odontogenic myxoma/myxofibroma. In: Barnes L, Eveson JW, Reichart P, Sidransky D, editors. Head and neck tumours. Pathology and genetics. WHO Classification of tumours. Lyon: IARC Press; 2005. p. 316.

not necessary for the diagnosis and are identical by trans-

- ² Buchner A, Merrell PW, Carpenter WM. Relative frequency of central odontogenic tumours: a study of 1,088 cases from Northern California and comparison to studies from other parts of the world. J Oral Maxillofac Surg 2006;64:1343-52.
- ³ Adebayo ET, Ajike SO, Adekeye EO. A review of 318 odontogenic tumours in Kaduna, Nigeria. J Oral Maxillofac Surg 2005;63:811-9.
- ⁴ Ladende AL, Ajayi OF, Ogunlewe MO, Adeyemo WL, Arotiba GT, Bamgbose BO, et al. *Odontogenic tumours: a review of 319 cases in a Nigerian teaching hospital*. Oral Surg Oral Med Oral Pathol Oral Radiol Endod 2005;99:191-5.
- ⁵ Ajavi OF, Ladende AL, Adeyemo WL, Ogunlewe MO. Odontogenic tumours in Nigerian children and adolescents. A retrospective study of 92 cases. World J Surg Oncol 2004;2:39-43.
- ⁶ Simon EN, Merkx MA, Vuhahula E, Ngassapa D, Stoelinga PJ. *Odontogenic myxoma: a clinicopathologic study of 33 cases*. Int J Oral Maxillofac Surg 2004;33:333-7.
- ⁷ Tamme T, Soots M, Kulla A, Karu K, Hanstein SM, Sokk A, et al. *Odontogenic tumours, a collaborative retrospective study of 75 cases covering more than 25 years from Estonia*. J Craniomaxillofac Surg 2004;32:161-5.
- Ochsenius S, Ortega A, Godoy L, Penafiel C, Escobar E. Odontogenic tumours in Chile: a study of 362 cases. J Oral Pathol Med 2002;31:415-20.

- Mosqueda-Taylor A, Ledesma-Montes C, Caballero-Sandoval S, Portilla-Robertson J, Ruiz-Godoy Rivera LM, et al. Odontogenic tumours in Mexico: a collaborative retrospective study of 349 cases. Oral Surg Oral Med Oral Pathol Oral Radiol Endod 1997;84:672-5.
- Barnes L. Tumours and tumour-like lesions of the soft tissues. In: Barnes L, editor. Surgical pathology of the head and neck. New York: Dekker 2001. p. 948-54.
- Slootweg PJ, Wittkampf ARM. Myxoma of the jaws. An analysis of 15 cases. J Max-Fac Surg 1986;14:46-52.
- Bast BT, Pogrel MA, Regezi JA. The expression of apoptotic proteins and matrix metalloproteinases in odontogenic myxomas. J Oral Maxilofac Surg 2003;61:1463-6.
- ¹³ Jaeger M, Santos J, Domingues M, Ruano R, Araujo N, Caroli A, et al. A novel cell line that retains the morphological characteristics of the cells and matrix of odontogenic myxoma. J Oral Pathol Med 2000;29:129-38.
- ¹⁴ Boson WL, Gomez RS, Araujo L, Kalapothakis E, Friedman E, De Marco L. *Odontogenic myxomas are not associated with activating mutations of the Gs alpha gene*. Anticancer Res 1998;18:4415-7.
- Perdigao PF, Stergiopoulos SG, De Marco L, Matyakhina L, Boikos SA, Gomez RS. Immunohistochemical investigation of protein kinase A regulatory subunit type 1A (PRKA-R1A) in odontogenic myxomas. Genes Chromosomes Cancer 2005;44:204-11.
- Perrotti V, Rubini C, Fioroni M, Piattelli A. Soft tissue myxoma: report of an unusual case located on the gingiva. J Clin Periodontol 2006;33:76-8.
- ¹⁷ Takahashi H, Fujita S, Okabe H. Immunohistochemical

- investigation in odontogenic myxoma. J Oral Pathol Med 1991;20:114-9.
- ¹⁸ Slootweg PJ, Koole R, Hordijk GJ. The presence of p53 protein in relation to the Ki-67 as cellular proliferation marker in head and neck squamous cell carcinoma and adjacent dysplastic mucosa. Oral Oncol Eur J Cancer 1994;30:138-41.
- ¹⁹ Van Diest PJ, Brugal G, Baak JPA. Proliferation markers in tumours: interpretation and clinical value. J Clin Pathol 1998;51:716-24.
- ²⁰ Lu QL, Abel P, Foster CS, Lalani EN. Bcl-2 role in epithelial differentiation and oncogenesis. Hum Pathol 1996;27:102-10.
- ²¹ Sampaio-Goes FC, Oliveira DT, Dorta RG, Nomogaki S, Landman G, Nishimoto IN, et al. Expression of PCNA, p53, Bax, and Bcl-X in oral poorly differentiated and basaloid squamous cell carcinoma: relationship with prognosis. Head Neck 2005;27:982-9.
- ²² Ribeiro DA, Salvadori DM, Marques ME. Abnormal expression of Bcl-2 and bax in rat tongue mucosa during the development of squamous cell carcinoma induced by 4-nitroquinoline 1-oxide. Int J Exp Pathol 2005;86:375-81.
- Martinez A, Brethauer U, Rojas IG, Spencer M, Mucientes F, Borlando J, et al. *Expression of apoptotic and cell proliferation regulatory proteins in actinic cheilitis*. J Oral Pathol Med 2005;34:257-62.
- ²⁴ Papa F, Scacco S, Vergari R, De Benedittis M, Petruzzi M, Lo Muzio L, et al. Expression and subcellular distribution of Bcl-2 and Bax proteins in serum-starved human keratinocytes and mouth carcinoma epidermoid cultures. Life Sci 2003;73:2865-72.
- ²⁵ Luo HY, Yu SF, Li TJ. Differential expression of apoptosisrelated proteins in various cellular components of ameloblastomas. Int J Oral Maxillofac Surg 2006;35:750-5.
- Lo Muzio L, Falaschini S, Farina A, Rubini C, Pezzetti F, Campisi G, et al. Bcl-2 as prognostic factor in head and neck squamous cell carcinoma. Oncol Res 2005;15:249-55.
- ²⁷ Loro LL, Johannessen AC, Vintermyr OK. Loss of Bcl-2 in the progression of oral cancer is not attributable to mutations. J Clin Pathol 1995;58:1157-62.
- ²⁸ Loyola AM, Cardoso SV, Lisa GS, Oliveira LJ, Mesquita RA, Carmo MA, et al. *Apoptosis in epithelial cells of apical radicular cysts*. Int Endod J 2005;38:465-9.
- ²⁹ Suzuki T, Kumamoto H, Kunimori K, Oooya K. *Immunohistochemical analysis of apoptosis-related factors in lining epithelium of radicular cysts*. J Oral Pathol Med 2005;34:46-52.
- ³⁰ Aoki T, Tsukinoki K, Karakida K, Ota Y, Otsuru M, Kaneko A. Expression of cyclooxygenase-2, Bcl-2 and Ki-67 in pleomorphic adenoma with special reference to tumour proliferation and apoptosis. Oral Oncology 2004;40:954-9.
- Takeda T, Sugihara K, Hirayama Y, Hirano M, Tanuma JI, Semba I. *Immunohistological evaluation of Ki67, p63, CK19 and p53 expression in oral epithelial dysplasias*. J Oral Pathol Med 2006;35:369-75.
- ³² Raju B, Mehrotra R, Oijordsbakken G, Al-Sharabi AK, Vasstrad EN, Ibrahim SO. Expression of p53, cyclin D1 and Ki-67 in pre-malignant oral lesions: association with clinicopathological parameters. Anticancer Res 2005;25:4699-706.
- ³³ Kurokawa H, Zhang M, Matsumoto S, Yamashita Y, Tanaka T, Tomoyose T, et al. *The relationship of the histologic grade at the deep invasive front and the expression of Ki-67 antigen and p53 protein in oral squamous carcinoma*. J Oral Pathol Med 2005;34:602-7.
- ³⁴ Van Heerden WF, Raubenheimer EJ, Dreyer L. The role of DNA ploidy and Ki-67 in the grading of mucoepidermoid carcinomas. Anticancer Res 2005;25:2589-92.
- 35 Saracoglu U, Kurt B, Gunham O, Given O. MIB-1 expression in odontogenic epithelial rests, epithelium of healthy oral mucosa and epithelium of selected odontogenic cysts. An immunohistochemical study. Int J Oral Maxillofac Surg 2005;34:432-5.

- ³⁶ Tosios KI, Kakarantza-angelopoulou E, Kapranos N. Immunohistochemical study of Bcl-2 protein, Ki-67 antigen and p53 protein in epithelium of glandular odontogenic cysts and dentigerous cysts. J Oral Pathol Med 2000;29:139-44.
- ³⁷ Birchall MA, Winterford CM, Allan DJ, Harmon BV. Apoptosis in normal epithelium, premalignant and malignant lesions of the oropharynx and oral cavity: a preliminary study. Oral Oncol, Eur J Cancer 1995;31B:380-3.
- ³⁸ Birchall MA, Schock E, Harmon BV, Gobé G. Apoptosis, mitosis, PCNA and bcl-2 in normal, leukoplakic and malignant epithelia of the human oral cavity. Oral Oncology 1997;33:419-25.
- Murti PR, Warnakulasuriya KA, Johnson NW, Bhonsle RB, Gupta PC, Daftary DK, et al. p53 expression in oral precancer as a marker for malignant potential. J Oral Pathol Med 1998;27:191-6.
- ⁴⁰ Ravi D, Nalinakumari KR, Rajaray RS, Krishnan Nair M, Radhakishna Pillai M. Expression of programmed cell death regulatory p53 and bcl-2 proteins in oral lesions. Cancer Lett 1996;105:139-46.
- Al Raybaud-Diogène H, Tétu B, Morenly R, Fortin A, Monteil RA. p53 overexpression in head and neck squamous cell carcinoma: review of the literature. Oral Oncol Eur J Cancer 1996;32B:143-9.
- ⁴² Bongers V, Snow GB, Van der Waal I, Braakhuis BJM. Value of p53 expression in oral cancer and adjacent normal mucosa in relation to the occurrence of multiple primary carcinomas. Oral Oncol, Eur J Cancer 1995;31B:392-5.
- ⁴³ Cruz IB, Snijders PJF, Meijer CJ, Braakhuis BJ, Snow GB, Wakboomers JM, et al. p53 expression above the basal cell layer in oral mucosa is an early event of malignant transformation and has predictive value for developing oral squamous cell carcinoma. J Pathol 1998;184:360-8.
- ⁴⁴ Jordan RCK, Catzavelos GC, Barrett AW, Speight PM. Differential expression of bcl-2 and Bax in squamous cell carcinoma of the oral cavity. Oral Oncol, Eur J Cancer 1996;32:394-400.
- ⁴⁵ Tsuji T, Mimura Y, Wen S, Li X, Kanekawa A, Sasaki K, et al. *The significance of PCNA and p53 protein in some oral tumours*. Int J Oral Maxillofac Surg 1995;24:221-5.
- ⁴⁶ Funaoka K, Arisue M, Kobayashi I, Iizuka T, Kohgo T, Amemiya A, et al. *Immunohistochemical detection of proliferating cell nuclear antigen (PCNA) in 23 cases of ameloblastoma*. Oral Oncol, Eur J Cancer 1996;32B:328-32.
- ⁴⁷ Kim J, In Yook J. *Immunohisthochemical study on proliferating cell nuclear expression in ameloblastomas*. Oral Oncol, Eur J Cancer 1994;30B:126-31.
- ⁴⁸ Tsai ST, Jin YT. Proliferating cell nuclear antigen (PCNA) expression in oral squamous cell carcinomas. J Oral Pathol Med 1995;24:313-5.
- ⁴⁹ Tsuji T, Shrestha P, Yamada K, Takagi H, Shinozaki F, Sasaki K, et al. Proliferating cell nuclear antigen in malignant and premalignant lesions of epithelial origin in the oral cavity and the skin: an immunohistochemical study. Virchows Archiv A Pathol Anat 1992;420:377-83.
- ⁵⁰ Girod SC, Pape HC, Krueger RF. p53 and PCNA expression in carcinogenesis of the oropharyngeal mucosa. Oral Oncol Eur J Cancer 1994;30B:419-23.
- Li TJ, Browne RM, Matthews JB. Expression of proliferating cell nuclear antigen (PCNA) and Ki-67 in unicystic ameloblastoma. Histopathology 1995;26:219-28.
- ⁵² Mighell A. *PCNA and p53*. Oral Oncol, Eur J Cancer 1995;31B:403-4.
- Li TJ, Browne RM, Matthews JB. Epithelial cell proliferation in odontogenic keratocysts. A comparative immunohistochemical study of Ki67 in simple, recurrent and basal cell naevus syndrome (BCNS)-associated lesions. J Oral Pathol Med 1995;24:221-6.
- ⁴ Tjalma W, Weyler J, Goovaerts G, De Pooter C, Van Marck E, Van Dam P. *Prognostic value of bcl-2 expression in pa-*

- tients with operable carcinoma of the uterine cervix. J Clin Pathol 1997;50:33-6.
- ⁵⁵ Ramsay JA, From L, Kahn HJ. Bcl-2 protein expression in melanocytic neoplasms of the skin. Mod Pathol 1995;8:150-4.
- Flohil CC, Janssen PA, Bosman FT. Expression of Bcl-2 protein in hyperplastic polyps, adenomas, and carcinomas of the colon. J Pathol 1996;178:393-7.
- ⁵⁶ Cho JH, Kim WH. Altered topographic expression of p21^{WAF1/} CIPI/SDII, Bcl-2 and p53 during gastric carcinogenesis. Pathol Res Pract 1998;194:309-17.
- De Angelis PM, Stokke T, Thorstensen L, Lothe RA, Clausen OPF. Apoptosis and expression of Bax, Bcl-x, and Bcl-2 apoptotic regulatory proteins in colorectal carcinomas, and associations with p53 genotype/phenotype. J Clin Pathol Mol Pathol 1998;51:254-61.
- ⁵⁹ Langlois NEI, Lamb J, Eremin O, Heys SD. Apoptosis in colorectal carcinoma occurring in patients aged 45 years and under: relationship to prognosis, mitosis, and immunohistochemical demonstration of p53, c-myc and bcl-2 protein products. J Pathol 1997;182:392-7.
- 60 Hindermann W, Berndt A, Wunderlich H, Katenkamp D, Kosmehl H. Quantitative evaluation of apoptosis and proliferation in renal cell carcinoma. Correlation to tumour subtype, cytological grade according to Thoenes-classification and the occurrence of metastasis. Pathol Res Pract 1997;193:1-7.
- ⁶¹ Barker BF. Odontogenic myxoma in El-Mofty SK Odontogenic tumours. Semin Diagn Pathol 1999;16:297-301.

- ⁶² Lombardi T, Luck C, Samson J, Odell EW. S100, alphasmooth muscle actin and cytokeratin 19 immunohistochemistry in odontogenic and soft tissue myxomas. J Clin Pathol 1995;48:759-62.
- ⁶³ Lo Muzio L, Nocini PF, Favia G, Procaccini M, Mignogna MD. Odontogenic myxoma of the jaws. A clinical, radiologic, immunohistochemical and ultrastructural study. Oral Surg Oral Med Oral Pathol Oral Radiol Endod 1996;82:426-33.
- 64 Schmidt-Westhausen A, Becker J, Schuppan D, Burkhardt A, Reichart PA. Odontogenic myxoma. Characterisation of the extracellular matrix (ECM) of the tumour stroma. Oral Oncol, Eur J Cancer 1994;30B:377-80.
- 65 Slootweg PJ, Van der Bos T, Straks W. Glycosaminoglycans in myxoma of the jaw: a biochemical study. J Oral Pathol 1985;14:299-306.
- ⁶⁶ Keszler A, Dominguez FV, Giannunzio G. Myxoma in childhood: an analysis of 10 cases. J Oral Maxillofac Surg 1995;53:518-21.
- ⁶⁷ Gundlach KH, Schulz A. Odontogenic myxoma. Clinical concept and morphological studies. J Oral Pathol 1977;6:343-58.
- ⁶⁸ Harrison JD. Odontogenic myxoma: ultrastructural and histochemical studies. J Clin Path 1973;26:570-82.
- ⁶⁹ White DK, Chen SY, Mohnac AM, Miller AS. Odontogenic myxoma. A clinical and ultrastructural study. Oral Surg Oral Med Oral Pathol 1975;39:901-17.
- ⁷⁰ Pahl S, Henn W, Binger T, Stein U, Remberger K. Malignant odontogenic myxoma of the maxilla: case with cytogenetic confirmation. J Laryngol Otol 2000;114:533-5.